

Review Article

Molecular pathogenesis and therapeutic strategies of Malignant melanoma: A brief Review

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ABSTRACT:

Melanoma is one of the most common and fastest growing cutaneous cancers worldwide. It contributes to 71–80 % of skin cancers deaths. UV radiation, melanotic nevi and sunlight are major risk factors leading to development of melanoma. These risk factors lead to various molecular changes allowing melanoma cells to have a growth and survival advantage over others. Depending on the features of the tumor (location, stage, and genetic profile), the therapeutic options may be surgical resection, chemotherapy, radiotherapy, photodynamic therapy (PDT), immunotherapy, or targeted therapy.

In this review, we discuss the various molecular changes associated with cutaneous melanoma and therapies approved or under evaluation for melanoma treatment and relevant research on the molecular mechanisms underlying melanogenesis.

Key words: Metastasis, UV radiation, Sunlight, mutation, BRAF, NRAS.

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INTRODUCTION

Cutaneous melanoma is a malignant neoplasm of the melanocytes which are found in the basal layer of the epidermis, the function of which is production of melanin; responsible for skin colour. It is one of the major types of skin cancer causing vast majority of skin cancer-related deaths even though it accounts for approximately 5% of skin cancer cases. (1, 2) There are four major forms of melanoma including (i) superficial spreading, (ii) nodular, (iii) lentigo maligna and (iv) acral lentiginous melanomas. Of these, the superficial spreading form remains most common; and accounts for about 70% of melanomas followed by nodular form that represents about 15–30% of melanoma cases.(3) Nowadays melanoma is considered as a multi-factorial disease which arises from an interaction between genetic susceptibility and environmental exposure and is known to be associated with various factors like UV radiation, excessive sun exposure, melanotic nevus which lead to its development. (4) Elwood *et al.* studied the correlation between melanoma and sun exposure concluding that intermittent sun exposure appears to be a major determinant of risk for melanoma and is highly

mutated tumour, characterised by an abundance of ultraviolet-induced DNA aberrations.(5) Some history of a sunburn may be an indicator of intense intermittent sun exposure. Moreover a history of sunburns in childhood is associated with the highest risk and sun exposure is believed to be involved in the malignant transformation of cutaneous melanocytes. (6,7) Primary melanoma patients demonstrate approximately an 11% mortality rate (2, 6), whereas the mortality rate due to metastatic melanoma is significantly higher and patients typically have a low survival rate due to the poor efficacies of current cancer therapies. (8,9) At the cellular level, cancer cells possess distinguishing molecular properties that allow for apoptosis evasion, limitless growth potential without the need for growth factors, angiogenesis, and metastasis. (10) Identifying specific molecular changes that allow melanoma cells to have a growth and survival advantage over others may aid in the development of more effective targeted therapies to improve the prognosis of melanoma patients.

In this review, we discuss the various molecular changes associated with cutaneous melanoma and therapies approved or under evaluation for melanoma

treatment and relevant research on the molecular mechanisms underlying melanogenesis.

Etiology and Pathogenesis

UV Radiation

Studies have shown that a major risk factor for melanoma development is exposure to Ultraviolet (UV) radiation. Excessive and cumulative exposure to UV radiation results in an increasing level of DNA damage to proliferating melanocytes, which eventually overcomes the capacity for DNA repair mechanisms to compensate usually evoking mechanisms which precipitate apoptotic cell death but occasionally a cell undergoes malignant transformation and nucleates a melanoma. (11,12) One or more blistering sunburn during childhood or adolescence doubles the risk for melanoma in later life and sunburns are known to be a measure of excessive sun exposure. (13) Ultraviolet radiation (UVR) is divided into ultraviolet C (UVC; 200 – 280 nm), ultraviolet B (UVB; 280 – 320 nm) and ultraviolet A (UVA; 320 – 400 nm). (14) The UVC spectrum is highly mutagenic but does not reach the earth's surface because it is absorbed by the stratospheric ozone layer meanwhile UVA and UVB wavelengths represent 95% and 5% of the UV spectrum reaching the earth's surface. (15-20) UVA is generally considered to be less carcinogenic than UVB, is an important factor involved in photocarcinogenesis acting through different mechanisms than UVB and low energy UVA radiation is weakly absorbed by DNA, but can be absorbed by other cellular chromophores, inducing oxidative changes in the cells. (21, 22) Meanwhile high-energy UVB photons are strongly attenuated by stratum corneum and in deeper layers of epidermis UV is absorbed by melanin, DNA, aminoacids, keratin, urocanic acid and other chromophores. Ultraviolet radiation also affects function of immune system causing immunosuppression is not limited only to irradiated area but a systemic suppression of immune system is observed. (24) The main cells affected by UV radiation are Langerhans cells and T lymphocytes which affects the number, function and morphology of Langerhans cells making them less capable of antigen presenting. The three molecular targets proposed to be involved in initiation of suppression of immune system are: DNA damage, plasma membrane damage and trans to cis isomerisation of urocanic acid, the latter affecting the morphology and function of Langerhans cells. (23, 25) Various experiments have shown evidence that UV radiation frequently leads to DNA mutations, such as formation of pyrimidine dimers or deamination of cytosine into thymidine. (26, 27)

Melanotic nevi

Nevi are benign lesions composed of a concentrated amount of melanocytes, leading to a dark spot on the skin due to large amount of melanin that is produced,

in one study about 81% of melanoma patients observed a changing nevus in the location of malignant lesion.(3, 28) The Clark model for melanoma progression emphasizes a series of histopathological changes beginning from benign melanocytic nevus to melanoma via dysplastic nevus. Various models of the genetic basis of melanoma development and progression are based on this Clark's multi-step model, predicting the acquisition of a BRAF mutation to be a founder event in melanocytic neoplasia. (29) Clark et al.(1984) also proposed a multi step progression model of melanoma. The first phenotypic change in normal melanocytes is the development of benign melanocytic nevus. Michaloglou et al. in 2008 suggested that melanocytic nevus is a benign clonal tumor, which temporarily undergoes proliferation via oncogenic BRAF signaling followed by growth arrest due to oncogene-induced senescence. (29) Dysplastic nevus, which histopathologically shows structural and cytological atypia is the next step towards the melanoma evolution. A dyspeptic nevus may arise from a preexisting melanocyte nevus or as a new lesion. The third step in progression is the radial growth phase (RGP) melanoma, which spreads progressively within or just beneath the epidermis followed by the final stage in melanoma progression which is the vertical growth phase (VGP), growing deep in the dermis and is a metastasis competent. (29) Mutations repressing apoptosis would be required for progression to VGP which allow cells to survive in the absence of keratinocytes. Miller and Mihm (2006) in a review stated that progression from RGP to VGP is also marked by the loss of E-cadherin, as well as the aberrant expression of N-cadherin and α V β 3 integrin. (29,30)

Molecular Changes Associated with Malignant Melanoma

BRAF

BRAF is the most frequently targeted gene in melanoma. BRAF gene is otherwise known as V-raf murine sarcoma virus oncogene homolog B1, is a member of the RAF family and is situated on chromosome 7q34. (31,32) It is a proto-oncogene that encodes a serine/threonine protein kinase as part of the RAS-RAF-MEK-ERK kinase pathway, which promotes cell growth and proliferation. (33,34) Studies have shown that around 40–60% of all melanoma cases exhibit an activated BRAF mutation occurring predominantly in melanomas arising on intermittently sun-exposed skin rather than in melanomas arising on chronic sun-exposed or completely unexposed sites. (35,36) Majority of BRAF mutations found in melanocytic nevi and melanomas substitute valine for glutamine at codon 600 which leads to a missense mutation at the kinase activation domain of exon 15. (37) More than 97% of BRAF mutations are located in codon 600 of the BRAF gene. (38) BRAF-mutated melanoma tends to

exhibit distinctive clinical features and is characterized by more aggressive biological behaviour and are more commonly found in younger patients and have tumors with superficial spreading or nodular histology and/or in anatomical regions without chronic sun damage. (39) Molecular testing for *BRAF* mutations in patients with advanced melanoma has become a standard for determining the course of therapy, and testing is recommended by the current National Comprehensive Cancer Network (NCCN) and European Society for Medical Oncology guidelines for melanoma. (40,41) Research has revealed that the early stage of melanoma formation, known as the radial growth phase, exhibits a low 10% *BRAF* mutation rate, which supports the hypothesis that *BRAF* does not play a part in melanoma initiation. (42) At the same time, 60–70% of vertical growth lesions and metastasized melanomas possess *BRAF* mutations, which suggests that this oncogenic mutation may be involved in cancer progression. (43–46) Thus identification of somatic mutations in the gene encoding serine-threonine protein kinase B-RAF (*BRAF*) in majority of melanomas gives an opportunity to test oncogene-targeted therapy for this disease.

NRAS

Another source of molecular changes that may allow for melanoma initiation or propagation is found in the *NRAS* GTPase or neuroblastoma RAS oncogene. (3) *NRAS* GTPase mutations are found in 15–20% of melanoma patients and activating mutations are usually found in the Q60/61 and G12/13 codons. (46,47) Activation of the MAP kinase pathway has been identified as a key player in melanoma. (48,49) Activating mutations in *NRAS* have been identified in approximately 15% – 20% of melanoma tumors. (50) The most common *NRAS* mutations are in exon 2 at codon 61, specifically Q61L (leucine substitution for glutamine) resulting in a constitutively active form of the protein leading to uncontrolled cellular proliferation. (51,52) The small GTPase, *NRAS*, was the first oncogene identified in melanoma and other mutational subtypes of melanoma. (52) Patients with mutant *NRAS* tumors tend to be older and have a history of chronic ultraviolet (UV) exposure. (53,54) Histologically, mutant *NRAS* tumors are more aggressive than other subtypes and have thicker lesions, elevated mitotic activity, and higher rates of lymph node metastasis. (55,56)

PI3K-ATK/PTEN

PI3K (phosphatidylinositol 3-kinase)–*AKT* pathway is a separate route that also plays a role in cell proliferation and survival. (57) Constitutive activation of *PI3K-ATK* leads to a competitive growth advantage that makes way for melanoma proliferation and metastasis. (58) High levels of *PI3K-ATK* activity can also result from activation via an *NRAS* mutation or a lack of inhibition due to *PTEN*

inactivation and *PTEN* containing a phosphatase domain, is inactivated in 12% of melanomas through mutation or methylation. (59, 60) Mutations of *PTEN* contribute to antiapoptosis, abnormal proliferation, angiogenesis, and invasion for melanoma development and progression. But analysis of melanoma tumor samples has identified about 3% *PI3K* missense mutation rate and such a low rate makes this pathway appear as an unlikely contributor to melanoma development and progressions. (61,62)

CDK4/CDKN2A

Cyclin-dependent kinase 4 (*CDK4*) classic cell cycle kinase forming complexes with D-type cyclins and Tumor suppressor gene *CDKN2A*, which encodes p16^{INK4a} have been implicated in familial melanoma development indicating the genetic susceptibility to develop melanoma. (63– 65) *CDK4* and p16^{INK4a} regulate cell cycle progression from the G1 to S phase upon activation by Cyclin D1. *CDK4* acts as a proto-oncogene that promotes the progression from G1 to S phase, allowing cell proliferation, whereas, p16^{INK4a} inhibits *CDK4* action, arresting cell division. (66) In view of these molecular changes, the logical step would be to inhibit *CDK4* to prevent further cell cycle progression and limit the uncontrolled growth in melanoma.

MC1R

Melanocortin 1 receptor (*MC1R*) acts as a G-protein coupled receptor in melanocytes located on chromosome 16q24.3 and plays a critical role in determining skin pigmentation. (3) *MC1R* is the most important gene found to play a role in predisposition to sporadic cutaneous melanoma and its association with cutaneous melanoma has been replicated and confirmed by meta-analyses and genome-wide association studies. (67) Exposure to UV radiation leads to generation of alpha-melanocyte stimulating hormone, which activates *MC1R* to generate melanin via the process of melanogenesis. (68) In normal conditions, melanin along with the glutathione pathway acts as antioxidants that neutralize the destructive effects of reactive oxygen species, however, in *MC1R* variant individuals, the decreased melanin production would lead to an increased state of oxidative stress and thus, DNA damage that may trigger mechanisms of melanoma initiation. (69) Individuals with *MC1R* variant leading to low melanin production are more vulnerable to UV-induced DNA mutagenesis making them susceptible to acquiring mutations in genetic loci of *BRAF*, *NRAS*, or *CDKN2A*, which have been found with constitutively activated *BRAF* melanoma patients who typically do not possess a history of chronic-sun damage of the skin. (68,70)

THERAPEUTICS STRATEGIES

Current therapeutic approaches include surgical resection, chemotherapy, photodynamic therapy

(PDT), immunotherapy, biochemotherapy, and targeted therapy. The therapeutic strategy includes single agents or combined therapies, depending on the patient's health, stage, and location of the tumor. (71) Primary melanomas are typically treated with surgical excision, yielding a high survival rate; but following metastasis, surgical excision of the tumor only yields about 10% five-year survival rate. (72) IFN- α is an FDA approved adjuvant therapy, which is administered after surgery in order to help prevent any remaining melanoma cells from proliferating. (73) Various treatment options available:

- **Chemotherapy:** It was the earliest treatment option for advanced melanoma. Chemotherapy combinations have been evaluated to improve the clinical responses, but the overall survival (OS) did not show improvement. (74) Dacarbazine, an alkylating agent approved in 1974 by FDA, is the standard chemotherapy medication for metastatic melanoma. Studies reported that a complete response was achieved in <5% and 5-year survival in 2%–6% of patients. (75) Electrochemotherapy is a technique that combines the use of cytotoxic drugs, bleomycin and cisplatin, with high-intensity electric pulses, which facilitates drug delivery into the cells. (76,77)
- **PDT:** It is a minimally invasive procedure requiring a photosensitizer and light of a defined wavelength, to activate the photosensitizer. (78) Both create reactive oxygen species (ROS) when combined with oxygen. ROS unleash irreversible damage to tumor cells and tumor-associated blood vessels, also activating antitumor, immune, and inflammatory responses. (79,80)
- **IMMUNOTHERAPY:** It is known in many types of cancer that complex interactions between the tumor and the immune system play a role in the metastatic spread to distant sites. (81) Tumor infiltrating lymphocytes (TILs) have been considered in many studies as independent markers for the occurrence of lymph node metastasis. TILs can mediate immune responses of the host against cancer cells, being associated with a positive outcome and improved survival in patients with malignant melanomas. (82) The immunogenic tumor microenvironment (TME), with mediators and cellular effectors of inflammation, influences the success of immunotherapies, the molecular pathways involved in this cancer-related inflammation are now being clarified, in order to establish new target molecules that may lead to improvements in the diagnosis and treatment of cancer. (83,84) Improved knowledge of the pathophysiology and a better understanding of the role of immune system in tumor evolution have led to the development and approval of several immunotherapies involving Interferon (IFN) α -

2b, Peginterferon α -2b (Peg-IFN), Interleukin-2 (IL-2), Treg inhibition, Cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) blockade, Programmed cell death protein 1 (PD-1)/PD-1 ligand (PD-L1) blockade, Oncolytic virus therapy, gp100 peptide vaccine, Adoptive T-cell therapy.

- **Targeted Therapy:** About 70% of patients with cutaneous melanoma harbor mutations in genes of key signaling pathways. These oncogenic mutations may be associated with melanoma cell proliferation and a malignant phenotype. (85) Targeted therapies include:

BRAF inhibitors

Vemurafenib: It is a highly selective inhibitor of mutated BRAF, is extremely active in patients with metastatic melanoma who harbor a mutation in *BRAFV600* (including patients with non V600E mutations) by inducing response rates in ~50% of patients and prolonging survival when compared to traditional chemotherapeutic agents. (86,87) Despite the clinical success of vemurafenib, most, if not all, patients eventually develop resistance. Multiple resistance mechanisms have been defined and are generally grouped as either those with reactivation of the MAPK pathway (intrinsic) or those outside of the MAPK pathway (extrinsic). (88) Vemurafenib is compared with dacarbazine for unresectable stage IIIC and stage IV metastatic *BRAFV600E* positive melanomas.

Dabrafenib: It is considered as a next generation agent and has a mechanism of action similar to that of vemurafenib. Dabrafenib is also a selective BRAF-mutant inhibitor approved by the FDA (2013) for the treatment of unresectable or metastatic melanomas harboring *BRAFV600E* mutations. (89,90)

MEK Inhibitors

Trametinib: It is a pharmacological MEK1/2 inhibitor with antitumoral activity, was approved (2013) as a monotherapy by the FDA for the treatment of unresectable or metastatic malignant melanomas with *BRAF* mutations. (91) Trametinib inhibits MEK i.e. the extracellular signal-regulated kinase that is downstream of BRAF, its recommended dose is 2 mg orally once daily and the duration of treatment remains until disease progression or unacceptable side effects develop. (3) Combined therapy of trametinib and dabrafenib (BRAF-mutant inhibitor) showed durable objective responses in a randomized, multicenter, open-label study and the combination was approved (2014) by the FDA for the treatment of unresectable and metastatic melanomas harboring *BRAF* mutations. (92,93)

CKIT Inhibitors:

Imatinib is an oral CKIT inhibitor that reveals significant activity in patients with metastatic

melanoma harboring *CKIT* aberrations, with a response rate of 30%. (94,95) Other multikinase inhibitors, such as sunitinib, dasatinib, and nilotinib, may have activity in patients with melanoma harboring *KIT* mutations. (90)

Cyclin-dependent kinase (CDK) inhibitors:

A new generation of selective CDK4/6 inhibitors, including ribociclib, abemaciclib, and palbociclib, has enabled tumors to be targeted with improved effectiveness and fewer adverse effects. (96) Abemaciclib has also been reported to induce growth regression in vemurafenib-resistant melanoma models, in which high levels of cyclin D1 expression and MAPK-pathway reactivation were observed. (97)

CONCLUSION

Melanoma represents a serious public health problem, due to its high case-fatality rate. Identification of individuals at high risk would be of major interest to improve early diagnosis and ultimately survival. The understanding of melanoma pathogenesis is crucial for the development of new therapeutic modalities. Characterization of oncogenic signaling pathways and interactions allow the identification of novel targets for clinically effective treatments, such as pathways inhibitors and immune checkpoint antibodies. Although it represents an advancement for melanomas treatment, these types of approaches face several challenges. The comprehensive features of patients that will benefit from each strategy aim to establish biomarkers (eg, specific mutations) for the best (eg, targeted) therapy in advanced melanomas. The convergence of immunology with other disciplines of biomedical research would lead to further improvement in the development of newer and more efficacious therapeutic approaches to manage this debilitating and deadly disease.

REFERENCES

1. Lens MB, Dawes M. Global perspectives of contemporary epidemiological trends of cutaneous malignant melanoma. *Br J Dermatol.* 2004;150:179–85.
2. Sboner A, Eccher C, Blanzieri E, et al. A multiple classifier system for early melanoma diagnosis. *Artif Intell Med.* 2003;27:29–44.
3. Liu Y, Sheikh M. Melanoma: Molecular Pathogenesis and Therapeutic Management, *Mol Cell Pharmacol.* 2014;6(3):228.
4. Rastrelli M, Tropea S, Rossi CR, Alaibac M, Melanoma: Epidemiology, risk factors, pathogenesis, diagnosis and classification, *In Vivo* November-December 2014 vol. 28 no. 6:1005-1011.
5. Sera F, Gandini S, Cattaruzza MS, Pasquini P, Picconi O, Boyle P, Melchi CF: Meta-analysis of risk factors for cutaneous melanoma: II. Sun exposure. *Eur J Cancer* 41: 45-60, 2015.
6. Elwood JM, Jopson J: Melanoma and sun exposure: an overview of published studies. *Int J Cancer* 73: 198-203, 1997.
7. Moan, J., Porojnicu, A.C., Dahlback, A.: Ultraviolet radiation and malignant melanoma. *Adv. Exp. Med. Biol.* 2008;624, 104–116
8. Thompson JF, Scolyer RA, Kefford RF. Cutaneous melanoma. *Lancet.* 2005;365:687–701.
9. Beddingfield FC., 3rd The melanoma epidemic: Res ipsa loquitur. *Oncologist.* 2003;8:459–65.

10. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell.* 2000;100:57–70.
11. Rass, K., Reichrath, J.: UV damage and DNA repair in malignant melanoma and nonmelanoma skin cancer. *Adv. Exp. Med. Biol.* 2008; 624, 162–178.
12. Kim, Y.G., Kim, H.J., Kim, D.S., Kim, S.D., Han, W.S., Kim, K.H., Chung, J.H., Park, K.C.: Up-Regulation and redistribution of Bax in ultraviolet B-irradiated melanocytes. *Pigment Cell Res.* 2000; 13, 352–357.
13. Weinstock MA, Colditz GA, Willett WC, et al. Nonfamilial cutaneous melanoma incidence in women associated with sun exposure before 20 years of age. *Pediatrics.* 1989;84:199–204.
14. Slominski A, Pawelek J. Animals under the sun: Effects of UV radiation on mammalian skin. *Clin Dermatol.* 1998;16:503–515.
15. Gambichler T, Breuckmann F, Boms S, Altmeyer P, Kreuter A. Narrowband UVB phototherapy in skin conditions beyond psoriasis. *J Am Acad Dermatol.* 2005;52(4):660–70.
16. Barbagallo J, Spann CT, Tutrone WD, Weinberg JM. Narrowband UVB phototherapy for the treatment of psoriasis: A review and update. *Cutis.* 2001;68(5):345–7.
17. Ledo E, Ledo A. Phototherapy, photochemotherapy, and photodynamic therapy: Unapproved uses or indications. *Clin Dermatol.* 2000;18(1):77–86.
18. Gasparro FP. The role of PUVA in the treatment of psoriasis. Photobiology issues related to skin cancer incidence. *Am J Clin Dermatol.* 2000;1(6):337–48.
19. Breuckmann F, Gambichler T, Altmeyer P, Kreuter A. UVA/UVA1 phototherapy and puva photochemotherapy in connective tissue diseases and related disorders: A research based review. *BMC Dermatol.* 2004;4(1)11.
20. Scheinfeld N, Deleo V. A review of studies that have utilized different combinations of psoralen and ultraviolet B phototherapy and ultraviolet A phototherapy. *Dermatol Online J.* 2003;9(5):7.
21. Agar NS, Halliday GM, Barnetson RS, Ananthaswamy HN, Wheeler M, Jones AM. The basal layer in human squamous tumors harbors more UVA than UVB fingerprint mutations: A role for UVA in human skin carcinogenesis. *Proc Natl Acad Sci U S A.* 2004;101(14):4954–9.
22. Slominski A, Pawelek J. Animals under the sun: Effects of UV radiation on mammalian skin. *Clin Dermatol.* 1998;16:503–515.
23. Anna B, Blazej Z, Jacqueline G, Andrew CJ, Jeffrey R, Andrzej S, Mechanism of UV-related carcinogenesis and its contribution to nevi/melanoma, *Expert Rev Dermatol.* 2007; 2(4): 451–469.
24. Norval M. The mechanisms and consequences of ultraviolet-induced immunosuppression. *Prog Biophys Mol Biol.* 2006;92(1):108–18.
25. Jhappan C, Noonan FP, Merlino G. Ultraviolet radiation and cutaneous malignant melanoma. *Oncogene.* 2003;22(20):3099–112.
26. Ikehata H, Ono T. The mechanisms of UV mutagenesis. *J Radiat Res.* 2011;52:115–25.
27. Agar N, Young AR. Melanogenesis: A photoprotective response to DNA damage, *Mutat Res.* 2005;571:121–32.
28. Rhodes AR, Weinstock MA, Fitzpatrick TB, Mihm MC, Jr, Sober AJ. Risk factors for cutaneous melanoma. A practical method of recognizing predisposed individuals. *JAMA.* 1987;258:3146–54.
29. Takata M, Murata H, Saida T. Molecular pathogenesis of malignant melanoma: a different perspective from the studies of melanocytic nevus and acral melanoma. *Pigment Cell Melanoma Res.* 23; 64–71.
30. Ackerman, A.B., and Mihara, I. (1985). Dysplasia, dysplastic melanocytes, dysplastic nevi, the dysplastic nevus syndrome, and the relation between dysplastic nevi and malignant melanomas. *Hum. Pathol.* 16, 87–91.
31. Jiang HW, Wortsman J, Matsuoka L, Granese J, Carlson JA, Mihm M, Slominski A. Molecular spectrum of pigmented skin lesions: from nevus to melanoma. *Expert Rev Dermatol.* 2006;1(5):679–700. 22.
32. Gray-Schopfer VC, Karasarides M, Hayward R, Marais R. Tumor necrosis factor-alpha blocks apoptosis in melanoma

- cells when BRAF signaling is inhibited. *Cancer Res.* 2007;67(1):122–9.
33. Kumar R, Angelini S, Czene K, et al. BRAF mutations in metastatic melanoma: A possible association with clinical outcome. *Clin Cancer Res.* 2003;9:3362–8.
 34. Houben R, Becker JC, Kappel A, et al. Constitutive activation of the ras-raf signaling pathway in metastatic melanoma is associated with poor prognosis. *J Carcinog.* 2004;3:6.
 35. Flaherty KT, Puzanov I, Kim KB, et al. Inhibition of mutated, activated BRAF in metastatic melanoma. *N Engl J Med.* 2010;363:809–19.
 36. Lang J, MacKie RM. Prevalence of exon 15 BRAF mutations in primary melanoma of the superficial spreading, nodular, acral, and lentigo maligna subtypes. *J Invest Dermatol.* 2005;125(3):575–9.
 37. Gill M, Celebi JT. B-RAF and melanocytic neoplasia. *J Am Acad Dermatol.* 2005;53(1):108–14.
 38. Ihle MA, Fassunke J, König K et al. Comparison of high resolution melting analysis, pyrosequencing, next generation sequencing and immunohistochemistry to conventional Sanger sequencing for the detection of p.V600E and non-p.V600E BRAF mutations. *BMC Cancer* 2014;14:13.
 39. Long GV, Menzies AM, Nagrial AM et al. Prognostic and clinicopathologic associations of oncogenic BRAF in metastatic melanoma. *J Clin Oncol* 2011;29:1239–1246.
 40. Dummer R, Hauschild A, Lindenblatt N et al. Cutaneous melanoma: ESMO clinical practice guidelines for diagnosis, treatment, and follow-up. *Ann Oncol* 2016;Suppl 5:126–132.
 41. Garbe C, Peris K, Hauschild A et al. Diagnosis and treatment of melanoma. European consensus-based interdisciplinary guideline-update 2016. *Eur J Cancer* 2016;63:201–217.
 42. Dong J, Phelps RG, Qiao R, Yao S, Benard O, Ronai Z, Aronson SA. BRAF oncogenic mutations correlate with progression rather than initiation of human melanoma. *Cancer Res.* 2003;63:3883–5.
 43. Pollock PM, Harper UL, Hansen KS, et al. High frequency of BRAF mutations in nevi. *Nat Genet.* 2003;33:19–20.
 44. Brose MS, Volpe P, Feldman M, et al. BRAF and RAS mutations in human lung cancer and melanoma. *Cancer Res.* 2002;62:6997–7000.
 45. Davies H, Bignell GR, Cox C, et al. Mutations of the BRAF gene in human cancer. *Nature.* 2002;417:949–54.
 46. Kelleher FC, McArthur GA. Targeting NRAS in melanoma. *Cancer J.* 2012;18:132–6.
 47. Davies MA. The role of the PI3K-AKT pathway in melanoma. *Cancer J.* 2012;18:142–7.
 48. Ball NJ, Yohn JJ, Morelli JG, et al. Ras mutations in human melanoma: a marker of malignant progression. *J Invest Dermatol.* 1994;102:285–90.
 49. Curtin JA, Fridlyand J, Kageshita T, et al. Distinct sets of genetic alterations in melanoma. *N Engl J Med.* 2005;353:2135–47.
 50. Edlundh-Rose E, Egyhazi S, Omholt K, et al. NRAS and BRAF mutations in melanoma tumours in relation to clinical characteristics. a study based on mutation screening by pyrosequencing. *Melanoma Res.* 2006;16:471–8.
 51. Russo AE, Torrisi E, Bevelacqua Y, et al. Melanoma: molecular pathogenesis and emerging target therapies (Review) *Int J Oncol.* 2009;34:1481–9.
 52. Couselo EM, Adelantado EZ, Ortiz C, García JS, García JP. NRAS-mutant melanoma: current challenges and future prospect. *OncoTargets and Therapy* 2017;10: 3947.
 53. Curtin JA, Fridlyand J, Kageshita T et al. Distinct sets of genetic alterations in melanoma. *N Engl J Med.* 2005;353(20):2135–2147.
 54. Jakob JA, Bassett RL Jr, Ng CS, et al. NRAS mutation status is an independent prognostic factor in metastatic melanoma. *Cancer.* 2012; 118(16):4014–4023.
 55. Thumar J, Shahbazian D, Aziz SA, Jilaveanu LB, Kluger HM. MEK targeting in N-RAS mutated metastatic melanoma. *Mol Cancer.* 2014;13:45.
 56. Devitt B, Liu W, Salemi R, et al. Clinical outcome and pathological features associated with NRAS mutation in cutaneous melanoma. *Pigment Cell Melanoma Res.* 2011;24(4):666–672.
 57. Fresno Vara JA, Casado E, de Castro J, Cejas P, Belda-Iniesta C, Gonzalez-Baron M. PI3K/akt signalling pathway and cancer. *Cancer Treat Rev.* 2004;30:193–204.
 58. Hennessy BT, Smith DL, Ram PT, Lu Y, Mills GB. Exploiting the PI3K/AKT pathway for cancer drug discovery. *Nat Rev Drug Discov.* 2005;4:988–1004.
 59. Davies MA. The role of the PI3K-AKT pathway in melanoma. *Cancer J.* 2012;18:142–7.
 60. Maehama T, Dixon JE. PTEN: a tumour suppressor that functions as a phospholipid phosphatase. *Trends in Cell Biology.* 1999;9(4):125–128.
 61. Curtin JA, Stark MS, Pinkel D, Hayward NK, Bastian BC. PI3-kinase subunits are infrequent somatic targets in melanoma. *J Invest Dermatol.* 2006;126:1660–3.
 62. Omholt K, Krockel D, Ringborg U, Hansson J. Mutations of PIK3CA are rare in cutaneous melanoma; *Melanoma Res.* 2006; 16: 197–200.
 63. Sherr CJ, Roberts JM. Living with or without cyclins and cyclin-dependent kinases. *Genes Dev.* 2004;18:2699–711. doi: 10.1101/gad.1256504.
 64. Tigan AS, Bellutti F, Kollmann K, Tebb G, Sexl V. CDK6-a review of the past and a glimpse into the future: from cell-cycle control to transcriptional regulation. *Oncogene.* 2016;35:3083–91. doi: 10.1038/ncr.2015.407.
 65. Liu L, Dilworth D, Gao L, et al. Mutation of the CDKN2A 5' UTR creates an aberrant initiation codon and predisposes to melanoma. *Nat Genet.* 1999;21:128–32.
 66. Bartkova J, Lukas J, Guldborg P, et al. The p16-cyclin D/Cdk4-pRb pathway as a functional unit frequently altered in melanoma pathogenesis. *Cancer Res.* 1996;56:5475–83.
 67. Tagliabue E, Gandini S, Bellocchio R, Maisonneuve P, Bishop JN, Polsky D, Lazovich D, Kanetsky P, Ghiorzo P, Gruis N, Landi MT, Menin C, Fargnoli M, García-Borrón JC, Sera F, Raimondi S, *MC1R* variants as melanoma risk factors independent of at-risk phenotypic characteristics: a pooled analysis from the M-SKIP project. *Cancer Manag Res.* 2018; 10: 1143–1154.
 68. Landi MT, Bauer J, Pfeiffer RM, et al. *MC1R* germline variants confer risk for BRAF-mutant melanoma. *Science.* 2006;313:521–2.
 69. Meyskens FL, Jr, Farmer P, Fruehauf JP. Redox regulation in human melanocytes and melanoma. *Pigment Cell Res.* 2001;14:148–54.
 70. Naysmith L, Waterston K, Ha T, et al. Quantitative measures of the effect of the melanocortin 1 receptor on human pigmentation status. *J Invest Dermatol.* 2004;122:423–8.
 71. Domingues B, Lopes JM, Soares P, Pópulo H. Melanoma treatment in review, *Immunotargets Ther.* 2018; 7: 35–49.
 72. Bhatia S, Tykodi SS, Thompson JA. Treatment of metastatic melanoma: An overview. *Oncology (Williston Park)* 2009;23:488–96.
 73. Parmiani G, Castelli C, Rivoltini L, et al. Immunotherapy of melanoma. *Semin Cancer Biol.* 2003;13:391–400.
 74. Wilson MA, Schuchter LM. *Melanoma.* New York: Springer; 2016. *Chemotherapy for melanoma*; pp. 209–229.
 75. Kim C, Lee CW, Kovacic L, Shah A, Klasa R, Savage KJ. Long-term survival in patients with metastatic melanoma treated with DTIC or temozolomide. *Oncologist.* 2010;15(7):765–771.
 76. Testori A, Ribero S, Bataille V. Diagnosis and treatment of in-transit melanoma metastases. *Eur J Surg Oncol.* 2017;43(3):544–560.
 77. Miklavčič D, Serša G, Breclj E, et al. Electrochemotherapy: technological advancements for efficient electroporation-based treatment of internal tumors. *Med Biol Eng Comput.* 2012;50(12):1213–1225
 78. Austin E, Mamalis A, Ho D, Jagdeo J. Laser and light-based therapy for cutaneous and soft-tissue metastases of malignant melanoma: a systematic review. *Arch Dermatol Res.* 2017;390(4):229–242.
 79. Dougherty TJ, Gomer CJ, Henderson BW, et al. Photodynamic therapy. *J Natl Cancer Inst.* 1998;90(12):889–905.
 80. Brown SB, Brown EA, Walker I. The present and future role of photodynamic therapy in cancer treatment. *Lancet Oncol.* 2004;5(8):497–508.

81. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. *CA Cancer J Clin.* 2009;59(4):225–249.
82. Gata VA, Lisencu CI, Vlad CI, Piciu D, Irimie A, Achimas-Cadariu P. Tumor infiltrating lymphocytes as a prognostic factor in malignant melanoma. Review of the literature. *J BUON.* 2017;22(3):592–598.
83. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature.* 2008;454(7203):436–444.
84. Gasser S, Lim LH, Cheung FS. The role of the tumour microenvironment in immune therapy. *Endocr Relat Cancer.* 2017;24(12):ERC-17-0146.
85. Flaherty KT. Targeting metastatic melanoma. *Annu Rev Med.* 2012;63:171–183.
86. Flaherty KT, Puzanov I, Kim KB, et al. Inhibition of mutated, activated BRAF in metastatic melanoma. *N Engl J Med.* 2010;363:809–819.
87. Sosman JA, Kim KB, Schuchter L, et al. Survival in BRAF V600-mutant advanced melanoma treated with vemurafenib. *N Engl J Med.* 2012;366:707–714.
88. Sumimoto H, Imabayashi F, Iwata T, Kawakami Y. The BRAF-MAPK signaling pathway is essential for cancer-immune evasion in human melanoma cells. *J Exp Med.* 2006;203:1651–1656.
89. Ballantyne AD, Garnock-Jones KP. Dabrafenib: first global approval. *Drugs.* 2013;73(12):1367–1376.
90. Livingstone E, Zimmer L, Vaubel J, Schadendorf D. BRAF, MEK and KIT inhibitors for melanoma: adverse events and their management. *Chin Clin Oncol.* 2014;3(3):29.
91. Wright CJ, McCormack PL. Trametinib: first global approval. *Drugs.* 2013;73(11):1245–1254.
92. Flaherty KT, Infante JR, Daud A, et al. Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations. *N Engl J Med.* 2012;367(18):1694–1703.
93. Niezgodna A, Niezgodna P, Czajkowski R. Novel approaches to treatment of advanced melanoma: a review on targeted therapy and immunotherapy. *Biomed Res Int.* 2015;2015:851387.
94. Hodi FS, Corless CL, Giobbie-Hurder A, et al. Imatinib for melanomas harboring mutationally activated or amplified KIT arising on mucosal, acral, and chronically sun-damaged skin. *J Clin Oncol.* 2013;31(26):3182–3190.
95. Guo J, Si L, Kong Y, et al. Phase II, open-label, single-arm trial of imatinib mesylate in patients with metastatic melanoma harboring c-Kit mutation or amplification. *J Clin Oncol.* 2011;29(21):2904–2909.
96. O’leary B, Finn RS, Turner NC. Treating cancer with selective CDK4/6 inhibitors. *Nat Rev Clin Oncol.* 2016;13(7):417–430.
97. Yadav V, Burke TF, Huber L, et al. The CDK4/6 inhibitor LY2835219 overcomes vemurafenib resistance resulting from MAPK reactivation and cyclin D1 upregulation. *Mol Cancer Ther.* 2014;13(10):2253–2263.