

Review Article

Periodontal Vaccine – A Review

Saikat Roy Chowdhury, Arijit Sarkar, Rana Praween Kumar, Vinod Kumar, Dipti Saxena, Bazro Deb Barman

Department of Periodontology, Awadh Dental College Jamshedpur, Jharkhand, India

ABSTRACT:

Vaccination is a process that induces specific immune resistance to a bacterial or viral infection. Edward Jenner developed and established the principle of vaccination using the cross protection conferred by cowpox virus, which is non pathogenic in humans. With the rapid growth of microbial genome sequencing and bioinformatics analysis tools we have the potential to examine all the genes and proteins from any human pathogen. This technique has the capability to provide us with new targets for anti-microbial drugs and vaccines. However, to realize this potential new bioinformatics and experimental approaches to select these targets from the myriad of available candidates are required.

Key words: Immune response, periodontal vaccine, periodontitis.

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Corresponding author: Dr. Saikat Roy Chowdhury, NH-33, Danga, Bhilai pahari, Jamshedpur-831012, Jharkhand, India

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INTRODUCTION

Periodontal diseases belong to a heterogeneous family of diseases, which demands a clear need for a better understanding of the etiology and pathogenesis behind formulation of a vaccine against the same. Both specific and nonspecific plaque hypothesis has its own merits and demerits(1,2).

Extensive research has been conducted to determine the role of cell-mediated immunity and serum antibodies in protection against infectious agents, less is known about the role of mucosal immunity(3).

Vaccination is a process that induces specific immune resistance to a bacterial or viral infection.

SPECIFIC IMMUNE RESPONSE

Chronic inflammation, if protracted, can result in an adaptation called the specific immune response. The specific immune response requires lymphocytes that use two types of receptors to generate specific immune responses, the b-cell antigen receptor and the t-cell antigen receptor.

Four phases are involved in the generation of specific immunity: [2]

- Clonal selection – Selection of lymphocytes that bear receptors recognizing the specific antigen

- Clonal expansion – Proliferation of those lymphocytes
 - Clonal contraction – Death of effector lymphocytes
 - Memory – Maintenance of an expanded clone of cells that bear the specific receptors recognizing the antigen.
- “Vaccination is the development of immunity or resistance to infection, after a secondary response (booster) that is adequate to consider the individual immune to a subsequent infection.”

Types of vaccination

Active immunization [3]: Here, an individual immune system is stimulated by administering killed or live attenuated products derived from micro-organisms.

Passive immunization [Figure 1]: Here, the antibodies formed in one individual are transferred to another.

DNA vaccination [Figure 2]: Here, DNA plasmids encoding genes required for antigen production are transferred to an individual.

Characteristics of an effective vaccine

- Safety
- Protectivity
- The ability to provide sustained protection
- The ability to produce neutralizing antibodies
- Stimulation of protective t-cells.

Practical considerations like

- Cost-effectiveness
- Biological stability
- Access
- Minimum contraindications and side effects

PATHOGENESIS OF PERIODONTITIS

Periodontitis is a disease of multifactorial origin with interaction among host, micro-organisms and environmental factors which includes genetic factors as well. Over 300 species of micro-organisms have been found to colonize the periodontal tissues, of which the following are considered to be the primary pathogens causing periodontitis [4-6]

- Porphyromonas gingivalis
- Aggregatibacter actinomycetemcomitans
- Tannerella forsythensis

These bacteria produce an array of antigens that stimulate pro-inflammatory cells and leads to the production of a wide variety of cytokines. These antigens may stimulate Th1 or Th2 cells.

Antigens are taken up by dendritic cells and presented to CD-8 or CD-4 cells along with MHC antigens. [7]

CD-8 cells → Th 1 response → CMI → Pro inflammatory
CD-4 cells → Th 2 response → Ab response → Protective

The host produces anti-bacterial substances such as defensins, cathelicidins and saposins, which protect the host tissues from bacterial products and forms the first line of defense. However, sometimes these are inactivated by the bacterial virulence factors. Once bacteria break this barrier, cytokines are produced, which can be both pro-inflammatory and anti-inflammatory. Production of inappropriate cytokines results in periodontitis. [7]

History of periodontal vaccines

From the time of Edward Jenner's discovery of small pox vaccine in 1796, antigens of infectious pathogenic bacteria and viruses have been the targets for a variety of vaccines against a number of infectious diseases. Thus, most vaccines target one or multiple antigenic components of mono-infecting bacteria or viruses. The principle of vaccination is based on two key elements of adaptive immunity namely specificity and memory. [3]

Three periodontal vaccines were employed 1870 Louis Pasteur creates 1st Live att. Bacterial vaccine (chicpenchoecra)

1885 Pasteur creates the first Live attenuated viral vaccine (rabies), 1886 Typhoid, 1900 Cholera, 1992 Hepatitis A, 1999 Meningococcal C Conjugate, 2004 DTaP/IPV DTa/IPV/HibTa/IPV, 2006 (Combine Hib) (Kudryar, et al.: Periodontal vaccine).

Mechanism of action

Types of periodontal immunization.

Active immunization

- Whole bacterial cells
- Sub unit vaccines
- Synthetic peptides as antigens

Passive immunization

- Murine monoclonal antibody
- Plantibodies

Genetic immunization

- Plasmid vaccines
- Live, viral vector vaccines

Active immunization

Here, the entire cell with its components is inoculated into a host to bring about active immunization.

• Klausen; 1991 [8] have shown that levels of serum antibody to both whole cells and partially purified fimbriae from *P. gingivalis* were elevated in rats immunized with *P. gingivalis* cells and that the activities of collagenase and cysteine proteinases in gingival and periodontal tissues were decreased.

• Kesavalu; 1992 [9] observed protection against invasion, but no colonization against *P. gingivalis* in a mouse chamber model by immunization with either killed heterologous invasive or non-invasive *P. gingivalis* strains. The immuneresponse to whole cells or selected envelope component did not completely abrogate lesions, but eliminated mortality.

Passive immunization

Passive immunization is short lived, because the host does not respond to the immunization and protection lasts only as long as the injected antibody persists. Here, the antigens are injected into a vector that produces antibodies.

These antibodies, when inoculated into a host, bring about passive immunization. Passive immunization can be brought about in two ways:

- Murine monoclonal antibodies
- Plantibodies

Genetic immunization

By the early 1990's, scientists had begun to study new approaches for the production of vaccines that differ in structure from traditional ones. The strategy involves genetic-engineering or recombinant DNA technology.

There are two types:

- Plasmid vaccines
- Live, viral vector vaccines

Preparations of human Periodontal Vaccine

Three types of vaccines were employed for the control of periodontal diseases. [31]

These include the vaccines prepared from:

- Pure cultures of streptococci and other oral organisms
- Autogenous vaccines, which are prepared from dental plaque samples of patients with destructive periodontal diseases. Plaque samples are removed from the diseased site and are sterilized by heat or by immersion in

iodine/formalin and are re-injected into the same patient, either locally or systemically.

• Stock vaccines such as Van Cott’s vaccine, Goldenberg’s vaccine, or Inava Endocorps vaccine.

Components of periodontal bacteria tested for antigenicity and potential as vaccine candidates

Generic name	Species name	Antigenic components
Porphyromonas	Intermedia	Whole cell non invasive 381 6235.2 (monkey isolate)
Porphyromonas	Macacae	Whole cell
Treponema	Denticola	Whole cell ATCC 35404
Fusobacterium	Nucleatum	Whole cell ATCC 25586
Actinobacillus	Actinomycete -mcomitans	Formalinized whole cell leucotoxin
Actinomyces	Viscosus	Fimbrial adhesins of T14V

Limitations of periodontal vaccines

However, several issues should be addressed pertinent to the development of a sophisticated vaccine against human periodontitis. Firstly, human periodontal disease is multifactorial caused by manifold pathogens. The intricacy of the periodontopathic bacteria might be a problem as a substantial number of bacteria appears to be involved in periodontal disease. The multiplicity of pathogenic organisms indicates that vaccine design against periodontitis is very complex.

Secondly, bacterial whole cells or crude extracts preparation for vaccination is not desirable because the antigenic determinants of bacteria potentially possess a high risk of cross reactivity with human counterparts.

Some more serious complication may stem from the vaccine or from the patient. Vaccines may be contaminated with unwanted proteins or toxins, or even live viruses. Supposedly killed vaccines may not have been properly killed; attenuated vaccines may revert to the wild type [3]. The patient may be hypersensitive to minute amounts of contaminating proteins, or immune-compromised, in which case any living vaccine is usually contraindicated.

Furthermore, importantly, animal models for vaccine trials may pose inconsistencies with human models in major histocompatibility complex-restriction of antigens presented by antigen presenting, thus obscuring the immunodominant epitope(s).

A humanized mouse system has been projected that has been reconstituted with human peripheral blood lymphocytes. This system needs to meet the requirement of least leakiness of a mouse immune system. More recently, a genetically engineered mouse system, such as the non-obese diabetes Non obese diabetic mouse CB 17- colony of BALB (mouse strain used in the study) prkdcscid/J mouse, has been initiated into the study of infectious and autoimmune

diseases in humans. This model may also prove to be a valuable tool for the study of periodontal disease and putative periodontal vaccines. [1]

As an innovative strategy, vaccines using cross reactive immunodominant epitopes as antigenic molecules in an attempt to stimulate antigen specific regulatory T-cells (Tregs, CD4+, CD25+, FoxP3+), secreting IL-10 and Transforming growth factor β , may provide new clues for periodontal disease prevention, through the induction of either immune tolerance or an effector function. [45]

Recently, a variety of strategies to enhance the immunogenicity of antigenic components of B or T lymphocytes have been adopted in vaccine trials against periodontal disease. These include, but not limited to, immunization of dendritic cells pulsed with antigens, the use of improved adjuvant formulas (e.g., the use of ofalum as an alternative to heat shock protein (Heat shock protein) based adjuvant), the use of recombinant plant monoclonal antibodies (plantibodies), [41,46,47] and the use of transgenic microorganisms as antigen vectors. [48,49] These efforts leave challenging areas to be chased further in the search for a more refined design that may guarantee the efficiency and safety of extended immune memory.

Future of periodontal vaccine-

As yet, there are no periodontal vaccine trials that have been successful in satisfying all requirements; to prevent the colonization of multiple pathogen biofilm in the subgingival area, to elicit a high level of effector molecules such as immunoglobulin sufficient to opsonize and phagocytose the invading organisms, to suppress alveolar bone loss, and to stimulate helper T-cell polarization that exerts cytokine functions optimal for protection against bacteria and tissue destruction.

As an innovative strategy, vaccines using cross-reactive immunodominant epitopes as antigenic molecules in an attempt to stimulate antigen-specific regulatory T-cells (Tregs, CD4+, CD25+, FoxP3+), secreting IL-10 and TGF- β , may provide new clues for periodontal disease prevention, through the induction of either immune tolerance or an effector function.

Periodontal disease as a multifactorial and polymicrobial disease requires a sophisticated vaccine design regimen targeting multiple pathogenic species. Vaccine regimens including the commonly shared antigens by selected periodontopathogenic species would be considered an innovative strategy.

Traditional periodontal vaccine trials aim to stimulate the immune system to produce increased levels of immunoglobulin of desired specificity. To accomplish this end, a conjugate vaccine (i.e. protein-CPS conjugate), dendritic-cell based immunotherapy, and subunit DNA vaccine encoding the desired immunogenic epitope have been devised.

Animal models for vaccine trials may pose discrepancies with human models in major histocompatibility complex-

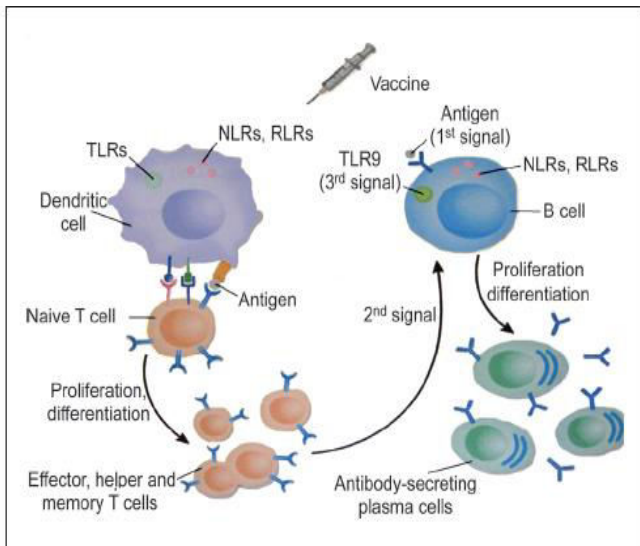
restriction of antigens presented by antigen presenting, thus obscuring the immunodominant epitope(s). A humanized mouse system has been proposed that has been reconstituted with human PBLs. This system needs to meet the requirement of least leakiness of a mouse immune system. More recently, a genetically engineered mouse system, such as the NOD.CB17-prkdc^{scid}/J mouse, has been introduced for the study of infectious and autoimmune diseases in humans. This model may also prove useful for the study of periodontal disease and putative periodontal vaccines.

CONCLUSION-

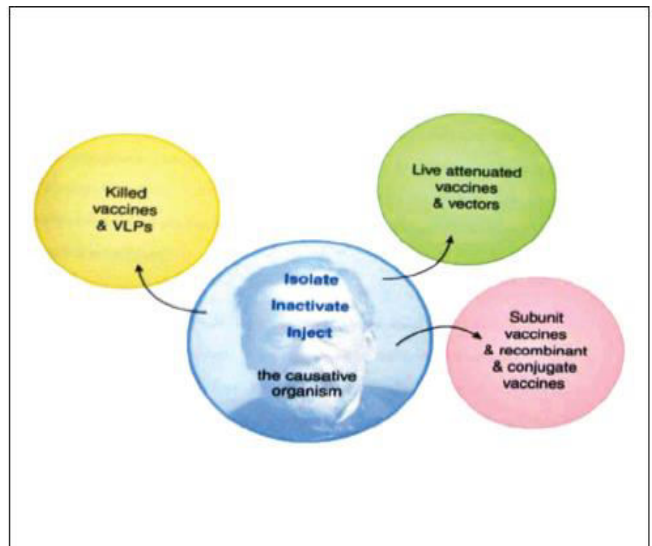
The current treatment of periodontitis is nonspecific and is centered on the removal of plaque by mechanical

debridement, often involving surgical procedures. This ongoing therapy is costly, painful and has a variable prognosis due in part to poor patient compliance.

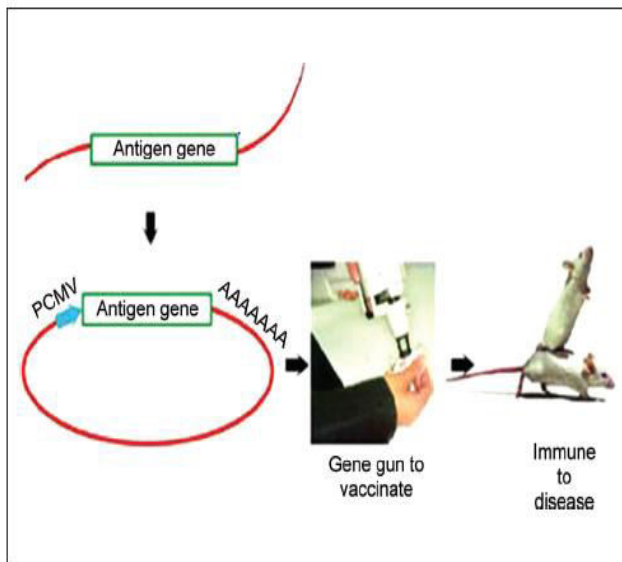
The use of antibiotics is limited by the need for constant treatment to prevent re-establishment of the pathogen. The elucidation of specific bacterial etiology suggests that the development of a specific treatment modality to target site colonization is now a rational approach to treat the disease. Vaccination may be an important adjunctive therapy to mechanical debridement in near future. It's not a myth but a reality which will come true in the near future if research is carried out in the right way in the right direction.



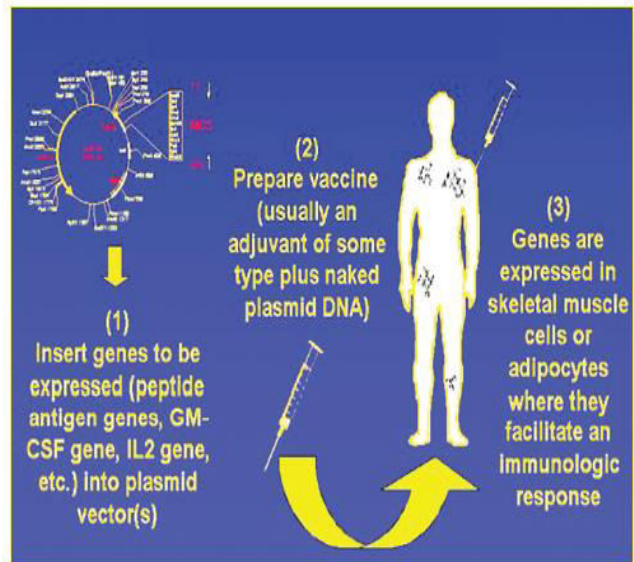
Mechanism of action



Active immunization



Genetic immunization



DNA vaccines

REFERENCES

1. Loesche WJ. Chemotherapy of dental plaque infections. *Oral Sci Rev* 1976;9:65.
2. Loesche WJ. Ecology of the oral flora. In: Nisengard RJ, Newman MG (Eds). *Oral microbiology and immunology*. Philadelphia: Saunders 1988:307.
3. Oral Immunization Using Live Attenuated Salmonella spp.as Carriers of Foreign Antigens. *Journal Clinical Microbiology Review* 1992.
4. Raghavendra Reddy, Yuvraja, Esther Nalini, Renuka Devi, Arun. Immunization in Periodontics: A Systematic Review. N
5. Verma JN, Rao M, Amselem S, Krzych U, Alving CR, Green SJ, et al. Adjuvant effects of liposomes containing lipid A: Enhancement of liposomal antigen presentation and recruitment of macrophages. *Infect Immun* 1992;60:2438-44.
6. Kudyar N, Dani N, Mahale S. Periodontal vaccine: A dream or reality. *J Indian SocPeriodontol* 2011;15:115-20.
7. McArthur WP, Magnusson I, Marks RG, Clark WB. Modulation of colonization by black pigmented Bacteroides species in squirrel monkeys by immunization with Bacteroides gingivalis. *Infect Immun* 1989;57:2313-7.
8. Persson GR, Engel LD, Whitney CW, Weinberg A, Moncla BJ, Darveau RP, et al. *Macaca fascicularis* as a model in which to assess the safety and efficacy of a vaccine for periodontitis. *Oral Microbiol Immunol* 1994;9:104-11.
9. Page RC, Schroeder HE. *Periodontitis in Man and Other Animals*. Basel: Karger; 1982.
10. Klausen B. Microbiological and immunological aspects of experimental periodontal disease in rats: A review article. *J Periodontol* 1991;62:59-73.
11. Kesavalu L, Ebersole JL, Machen RL, Holt SC. *Porphyromonas gingivalis* virulence in mice: Induction of immunity to bacterial components. *Infect Immun* 1992;60:1455-64.
12. Socransky SS, Haffajee AD. The bacterial etiology of destructive periodontal disease. *J Periodontol* 1992;63:322-31.
13. Harano K, Yamanaka A, Okuda K. An antiserum to a synthetic fimbrial peptide of *Actinobacillus actinomycetemcomitans* blocked adhesion of the microorganism. *FEMS Microbiol Lett* 1995;130:279-85.
14. Takamatsu Matsushita N, Yamaguchi N, Kawasaki M, Yamashita Y, Takehara T, Koga T. Immunogenicity of *Actinobacillus actinomycetemcomitans* serotype b specific polysaccharide protein conjugate. *Oral Microbiol Immunol* 1996;11:220-5.
15. Herminajeng E, Asmara W, Yuswanto A, Barid I, Sosroseno W. Protective humoral immunity induced by surface-associated material from *Actinobacillus actinomycetemcomitans* in mice.
16. Kobayashi T, Tahara T, Abiko Y, H Yoshie. Human monoclonal antibody as a periodontal vaccine. *J clin Periodontology suppl* 2003;4:11-2.
17. Bainbridge BW, Page RC, Darveau RP. Serum antibodies to *Porphyromonas gingivalis* block the prostaglandin E2 response to lipopolysaccharide by mononuclear cells. *Infect Immun* 1997;65:4801-5.
18. Gemmell E, Bird PS, Ford PJ, Ashman RB, Gosling P, Hu Y, et al. Modulation of the antibody response by *Porphyromonas gingivalis* and *Fusobacterium nucleatum* in a mouse model. *Oral Microbiol Immunol* 2004;19:247-51.
19. Choi JI, Borrello MA, Smith ES, Zauderer M. Polarization of *Porphyromonas gingivalis* specific helper T cell subsets by prior immunization with *Fusobacterium nucleatum*. *Oral Microbiol Immunol* 2000;15:181-7.
20. Choi J, Borrello MA, Smith E, Cutler CW, Sojar H, Zauderer M. Prior exposure of mice to *Fusobacterium nucleatum* modulates host response to *Porphyromonas gingivalis*. *Oral Microbiol Immunol* 2001;16:338-44.
21. Choi JI, Kim US, Kim SJ, Son WS, Park HR. *Fusobacterium nucleatum* impairs serum binding to *Porphyromonas gingivalis* biofilm. *Oral Microbiol Immunol* 2003;18:92-4.
22. Sjöström K, Darveau R, Page R, Whitney C, Engel D. Oposonic antibody activity against *Actinobacillus actinomycetemcomitans* in patients with rapidly progressive periodontitis. *Infect Immun* 1992;60:4819-25.
23. Giardino A, Ebersole JL, Holt SC. Characteristics of systemic antibody responses of nonhuman primates following active immunization with *Porphyromonas gingivalis*, *Prevotella intermedia* and *Bacteroides fragilis*. *Oral Microbiol Immunol* 1996;11:79-87.
24. Kohler JJ, Pathangey LB, Brown TA. Oral immunization with recombinant *Salmonella typhimurium* expressing a cloned *Porphyromonas gingivalis* hemagglutinin: Effect of boosting on mucosal, systemic and immunoglobulin G subclass response. *Oral Microbiol Immunol* 1998;13:81-8.
25. Kato H, Saito S, Takiguchi H, Abiko Y. Bactericidal activity of a monoclonal antibody against a recombinant 40 kDa outer membrane protein of *Porphyromonas gingivalis*. *J Periodontol* 2000;71:368-75.
26. Fan Q, Sims T, Sojar H, Genco R, Page RC. Fimbriae of *Porphyromonas gingivalis* induce oposonic antibodies that significantly enhance phagocytosis and killing by human polymorphonuclear leukocytes. *Oral Microbiol Immunol* 2001;16:144-52.
27. Persson GR, Engel D, Whitney C, Darveau R, Weinberg A, Brunsvold M, et al. Immunization against *Porphyromonas gingivalis* inhibits progression of experimental periodontitis in nonhuman primates. *Infect Immun* 1994;62:1026-31.
28. Nagasawa T, Aramaki M, Takamatsu N, Koseki T, Kobayashi H, Ishikawa I. Oral administration of *Porphyromonas gingivalis* fimbriae with cholera toxin induces anti fimbriae serum IgG, IgM, IgA and salivary IgA antibodies. *J Periodontal Res* 1999;34:169-74.
29. Chu L, Bramanti TE, Ebersole JL, Holt SC. Hemolytic activity in the periodontal pathogen *Porphyromonas gingivalis*: Kinetics of enzyme release and localization. *Infect Immun* 1991;59:1932-40.
30. Booth V, Ashley FP, Lehner T. Passive immunization with monoclonal antibodies against *Porphyromonas gingivalis* in patients with periodontitis. *Infect Immun* 1996;64:422-7.
31. Malhotra R, Kapoor A, Grover V, Tuli AK. Periodontal vaccine. *Indian J Dent Res* 2011;22:698-705.
32. Lee JY, Yi NN, Kim US, Choi JS, Kim SJ, Choi JI. *Porphyromonas gingivalis* heat shock protein vaccine reduces the alveolar bone loss induced by multiple periodontal pathogenic bacteria. *J Periodontal Res* 2006;41:10-4.
33. Curtis MA, Aduse Opoku J, Slaney JM, Rangarajan M, Booth V, Cridland J, et al. Characterization of an adherence and antigenic determinant of the ArgI protease of *Porphyromonas gingivalis* which is present on multiple gene products. *Infect Immun* 1996;64:2532-9.

34. Holt SC, Kesavalu L, Walker S, Genco CA. Virulence factors of *Porphyromonas gingivalis*. *Periodontol* 2000 1999;20:168 - 238.
35. Marawar PP, Devkar N. Gingipains: The virulence factor of *P. gingivalis*. *J Indian Soc Periodontol* 2004;7:95-9.
36. Kadowaki T, Yoneda M, Okamoto K, Maeda K, Yamamoto K. Purification and characterization of a novel arginine specific cysteine proteinase (argingipain) involved in the pathogenesis of periodontal disease from the culture supernatant of *Porphyromonas gingivalis*. *J Biol Chem* 1994;269:21371-8.
37. Imamura T. The role of gingipains in the pathogenesis of periodontal disease. *J Periodontol* 2003;74:111-8.
38. Moritz AJ, Cappelli D, Lantz MS, Holt SC, Ebersole JL. Immunization with *Porphyromonas gingivalis* cysteine protease: Effects on experimental gingivitis and ligature induced periodontitis in *Macaca fascicularis*. *J Periodontol* 1998;69:686-97.
39. Lehner T, Ma JK, Walker P, Childerstone A, Todryk S, Kendall H, et al. T cell and B cell epitope mapping and construction of peptide vaccines. In: Genco RJ, Hamada S, Lehner T, McGhee JR, Mergenhagen S, editors. *Molecular Pathogenesis of Periodontal Disease*. Washington, DC: ASM Press; 1994. p. 279-92.
40. Lee JY, Sojar HT, Bedi GS, Genco RJ. Synthetic peptides analogous to the fimbriin sequence inhibit adherence of *Porphyromonas gingivalis*. *Infect Immun* 1992;60:1662-70.
41. Ma JK, Hiatt A, Hein M, Vine ND, Wang F, Stabila P, et al. Generation and assembly of secretory antibodies in plants. *Science* 1995;268:716-9.
42. Okuda K, Kato T, Naito Y, Takazoe I, Kikuchi Y, Nakamura T, et al. Protective efficacy of active and passive immunizations against experimental infection with *Bacteroides gingivalis* in ligated hamsters. *J Dent Res* 1988;67:807-11.
43. Ishikawa I, Nakashima K, Koseki T, Nagasawa T, Watanabe H, Arakawa S, et al. Induction of the immune response to periodontopathic bacteria and its role in the pathogenesis of periodontitis. *Periodontol* 2000 1997;14:79-111.
44. Persson GR. Immune responses and vaccination against periodontal infections. *J Clin Periodontol* 2005;32:39-53.
45. Belkaid Y. Regulatory T cells and infection: A dangerous necessity. *Nat Rev Immunol* 2007;7:875-88.
46. Shin EA, Lee JY, Kim TG, Park YK, Langridge WH. Synthesis and assembly of an adjuvanted *Porphyromonas gingivalis* fimbrial antigen fusion protein in plants. *Protein Expr Purif* 2006;47:99-109.
47. Sharma A, Sojar HT, Hruby DE, Kuramitsu HK, Genco RJ. Secretion of *Porphyromonas gingivalis* fimbriin polypeptides by recombinant *Streptococcus gordonii*. *Biochem Biophys Res Commun* 1997;238:313-6.
48. Sharma A, Honma K, Evans RT, Hruby DE, Genco RJ. Oral immunization with recombinant *Streptococcus gordonii* expressing *Porphyromonas gingivalis* Fim A domains. *Infect Immun* 2001;69:2928-34.

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